

ISSN Print: 2664-9926 ISSN Online: 2664-9934 Impact Factor: RJIF 5.45 IJBS 2023; 5(1): 209-214 www.biologyjournal.net Received: 13-04-2023 Accepted: 14-05-2023

#### Abrahem Sadek

Addis Ababa University College of Veterinary Medicine, Bishoftu Ethiopia, Ethiopia

#### Megarsa Bedasa

Addis Ababa University College of Veterinary Medicine, Bishoftu Ethiopia, Ethiopia

#### **Ephrem Shimelis**

Addis Ababa University College of Veterinary Medicine, Bishoftu Ethiopia, Ethiopia

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# Isolation of salmonella from goats slaughtered at export abattoirs in Bishoftu town

## Abrahem Sadek, Megarsa Bedasa and Ephrem Shimelis

#### DOI: https://dx.doi.org/10.33545/26649926.2023.v5.i1c.188

#### Abstract

A cross-sectional study was conducted between February, 2020 and March 2020 on apparently healthy slaughtered goats at Abyssnia export abattoirs located at Bishoftu towns to estimate the prevalence of salmonellosis from goat meat by the isolation and identification conventional tests. A total of 200 samples carcass swab (60), hand swab (40), knife swab (40) and faces (60) were collected from goats slaughtered at Abyssinia export abattoir. The samples were examined for the presence of Salmonella by the conventional techniques of Selective media and biochemical tests for isolation salmonella species. From the total of 200 animals examined, 5 (2.5%) were positive of which 4 (2%) were young and 1 (0.5%) were adult. Statistically significant difference (p=0.01) in the prevalence of Salmonella was observed between the age of goats. Of the total 200 samples examined from apparently healthy slaughtered goats, 5 (2.5%) samples were Salmonella positive. Salmonella was isolated from 1.5% carcass swab, 0% hand swab, 0.5% knife swab and 0.5% faces samples. However, there was significant difference between samples (P=0.01). The results of this study showed the potential risk of goats as sources of pathogen for humans in the study area. These findings stressed the need for implementation of preventing close contact of offal and carcass during evisceration.

Keywords: Abattoir, Abyssnia, carcass, goats, isolation, Salmonella

#### Introduction

Ethiopia consist a diversity of livestock population and possess the large livestock population from the African countries (IBC, 2007)<sup>[1]</sup>. The goat population is estimated to be around 30.20 million heads.in which 1, 128, 000 goats were estimated to be consumed domestically (CSA, 2017). Livestock productions plays an important role to smallholder farmers and the national economy of the country in generating income to farmers, creating job opportunities, ensuring food security, providing services, contributing to asset, social, cultural and environmental values, and sustain livelihoods (Behnke, 2010; Endalew B, Ayalew, 2016)<sup>[4,</sup> <sup>12]</sup>. However, the sector is affected by infectious diseases (Viral, bacterial, parasitic and fungal diseases), feed shortage and lack of veterinary service (Ganteby, 1991) <sup>[17]</sup>. From bacterial disease, food borne zoonotic pathogenic disease such as Salmonella an importantly affect both human and animals in developing countries including Ethiopia. Salmonellosis is one of the widest spread food borne zoonoses with the main cause of mortality and morbidity to humans and animals (D'Aoust, 1994)<sup>[9]</sup>. Salmonellosis is a disease or a group of diseases caused by a wide variety of Salmonella serovars in various hosts including goat (OIE Manual, 2006) which remain a serious problem with public health significance throughout the world (Tabaraie et al., 1994) [34].

Salmonella are Gram negative, bacillus shaped, non-spore forming, aerobic and facultatively anaerobic organisms belong under family Enterobacteriaceae, were most of them have peritrichous flagella that use for motility (Freeman, 1985; Gene, 2002; OIE Manual, 2006) <sup>[13, 16]</sup>. They have two species such as, *Salmonella enterica* and *Salmonella bongori*. Salmonella enteric has six subdivisions of *S. enterica* subspecies 1-6 exist with over 2500 serovars currently identified and several common serovars to human clinical infections (Dworkin *et al.*, 2006) <sup>[11]</sup>. The main forms of salmonellosis happen are in three forms which is peracute septicemia, acute enteritis, and chronic enteritis. Common clinical signs of salmonellosis are abortion, upset stomach, diarrhea, fever, and pain and also cramping in your belly.

Corresponding Author: Abrahem Sadek Addis Ababa University College of Veterinary Medicine, Bishoftu Ethiopia, Ethiopia Although, the clinical signs may vary from species to species (Blood *et al.*, 2003) <sup>[5]</sup>. They are facultative intracellular pathogens that cause disease in animals, they are commonly asymptomatic carriers. Salmonella was a highly concentrated intensive study on cattle and sheep; however, there is little published information on the conveying of Salmonella in goats, although goat meat was one of a source of Salmonella food poisoning in Ethiopia (Kadaka *et al.* 2000) <sup>[22]</sup>.

The diagnosed salmonellosis can be based on taking history and laboratory examination of the Salmonellosis using feces, carcass swab, feed and water (Carlton, 1998)<sup>[8]</sup>. However, identification salmonella is the most difficult at clinical sign because the bacteria may be intermittently and it may be sample taking is important for examine. However, for identification of the bacteria at least requires as five days for use of standard cultural techniques (House and Smith, 1998)<sup>[18]</sup>. Thus, cultural technical for isolation of salmonellosis was: Pre-enrichment, enrichment, plating out and different biochemical and serological test according to the set of standards (Pieskus, 2006)<sup>[27]</sup>.

The Ethiopia goat population was suffering from a contagious disease like Salmonellosis and Brucellosis. Salmonella is the main causes of meat contamination that can affect animal and human being (Pepin *et al.*, 1997; Sierra *et al.*, 1995) <sup>[28, 33]</sup>. The risk factors for the exposure of salmonellosis contamination of animal carcasses and fresh meat during slaughter and dressing by the faeces, the skin, water, air, intestinal contents, processing equipment and humans, and can be transferred to the carcass during skin removal and evisceration (Hansson *et al.*, 2000; Reid *et al.*, 2002) <sup>[19, 32]</sup>. Therefore, this study was designed to isolation of salmonella from different type of samples (knife swab, hand swab, carcass swab and fecal) collected from goats slaughtered in Abyssinia export abattoir.

# Materials and Methods

## Study area

The study was conduct at Abyssinia export abattoirs in Bishoftu town, East Showa zone of Oromia regional state. Bishoftu town was located at 9°N latitude and 40°E longitude at altitude of 1850 masl with the annual average temperature ranges of 12.3 °C to 27.7 °C with an overall average of 18.7 °C. In thus, area the climate and soil type are the same with those in the highland areas in Ethiopia. It got heavy rain in the season of June to September with annual rain fall of 866 mm and which is 84% got in that long rainy season. However, the are many livestock farms of beef and dairy cattle, small ruminant, poultry and equines and there many modern slaughter services (abattoirs) in in the city. From these abattoirs, Ethiopia is exporting millions of tons of meat of cattle, sheep and goats to Middle East countries and gain good external currency. Hence, this designed to undertake salmonella isolation from goats brought to Abyssinia Slaughtering Service House for slaughter.

#### Study design and animals

The cross-sectional study was conducted on isolation and identification of salmonella from apparently healthy goats slaughtered at Abyssinia Export abattoir service from February 2020 to March 2020. Conveniently samples were collected from 200 goats purchased from different zones of Oromia region (Arsi, Hararghe, Bale and Borena). The animal was grouped into two classes as young and adult based on dentition by Vatta,  $(2005)^{[36]}$  where the animal which have only one pair permanent of incisors teeth were considered as young (<1 year to 2 years) while the goats which have two or more pairs of permanent incisors teeth were considered as adult (>2 years).

#### Sample collection and transportation

Based on its accessibility of sample during the sample collection, the sample were collected from 200 goats provided for slaughtering at Abyssinia export abattoir. The samples were collected aseptically from goats during slaughtering operation. Thus, sample such as: feces, carcass swab, knife swab and hand swab were collected aseptically using sterile cotton swab for organ samples and put into sterile universal bottles with peptone water media for transportation according to the recommendation (Quinn and Markey, 2003) <sup>[29]</sup>. Each sample was collected from each slaughtered goat during operation. Because of fasting abattoir worker do not taken four sample at the same time from the same goat. All samples were labeled apparent identifying species of the animal, age and type of sample and sampling ID number. After putting the samples in an ice box with ice pack. The sample was transported immediately to AAU, CVMA veterinary public health laboratory for bacteriological processing.

#### Isolation salmonella

# Sample preparation and inoculation on Nonselective Pre-enrichment

The fresh feces sample of 10 g was broken in to pieces by using Mortar and pestle and homogenized by homogenizer before transferred to pre-enriched in 90 ml of buffered peptone water (BPW). also. The swab samples were transferred to pre-enrichment of buffered peptone water in the ratio of 1:9 BPW (HI Media M1494, Mubi, India). The both the Swab and fecal sample were incubated for 24 hr at 37 °C.

#### Culturing salmonella on selective Media

On the second day, 0.1 ml (100 uL) of the pre-enrichment broth will be transferred to 10 ml Rappaport-Vassiliadis soy peptone (RVS) and incubated at 37 °C overnight (18-24 hours). On the third day, the suspension culture of the Rappaport-Vassiliadis Soya Peptone broth (RVS) was taken a loop full of the suspension was inoculated on to xylose lysine deoxycholate International Journal of Microbiology (XLD) (HiMedia M031, Mubi, India) agars and incubated at 37° C for 24 hr. The incubation was extended to 48 hr for those who did not show any growth during the 24 hr incubation. Typical Salmonella colonies having a slightly transparent zone of reddish color and a black center was isolated and sub cultured on nutrient agar (Oxoid CM0003, Basingstoke, England) and various biochemical tests were conducted.

#### Testing isolate by different biochemical tests

After the targeted bacteria was isolated and identified by using the selective media (XLD), the biochemical tests were performed by using: triple sugar iron agar (TSI), Methyl red (MR), Voges Proskauer (VP) (Himedia M070I, Mumbai, India), tryptone broth (Himedia M463, Mumbai, India) for indole test and Citrate tests for further identification and confirmation of the salmonella. Paternally (Positive and

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negative the above media), the salmonella was identified on the biochemical fermentation on the five above median. This mean that the presence of salmonella was interpreted by color change, gas formation and formation of hydrogen sulfide on the above media. After the salmonella was identified, it classified into motile and non-motile groups based on its motility in Motility media.

#### Data management and analysis

Data collected in the study and the results of laboratory investigations were entered into Microsoft Excel, edited, coded after done according to annex 3 and analyzed by statistical methods using proper statistical analysis (STATA) version 13.0. Descriptive statistics was used to determine the prevalence of salmonellosis in the study area. A P-value less than 0.05 at 95% confidence interval was considered for significance.

#### Results

This study conducted to isolate salmonella species from goats slaughtered at Abyssinia export abattoir during February 2020 to March 2020 to isolate salmonella species. A total of 200 samples from Abyssinia export abattoirs (60 carcass swab, 40 hand swab, 40 knife swab and 60 feces) were collected and examined for the presence of salmonella species. Among these five salmonella species was isolated of showing characteristics salmonella., i.e., the colony appears as translucent with the black center and hydrogen sulfide production from thiosulfate is easily detected because colonies become dark or gloomy, due to the precipitation of ferric sulfide on XLD agar and positive on biochemical tests for salmonella and. The Salmonella were isolated from all sample types except hand swab. The prevalence varied with sample site and time of sampling. When compere the prevalence of salmonella by the sample sources, the salmonella was more prevalent on carcasses (1.5%). In case of age, it's more prevalent on young (2%) than adult.

**Prevalence of Salmonella from Different Sample Sources** Salmonella was isolated from the samples collected from Carcass swab 3 (1.5%), Hand swab 0(0%), Knife swab 1 (0.5%) and Feces 1 (0.5%) as it isolated and identified on the Selective media and Biochemical analysis of the sample. There was significant difference (p < 0.05) in the frequency of Salmonella isolation among the samples (Table 1).

**Table 1:** The prevalence of salmonella from different sample sources

Types of samples	Examined	Positive	Percentage in each sample	Percentage in total	$\chi^2$	p-value
Carcass swab	60	3	5%	1.5%	6.4431	0.011
Hand swab	40	0	0%	0%		
Knife swab	40	1	2.5%	0.5%		
Feces	60	1	1.67%	0.5%		
Total	200	5	2.5%	2.5%		

#### Prevalence of salmonella between the age of goats

The prevalence of the salmonella was high in young goat when it compared with that of adult goats (2% and 0.5%, respectively) (Table 2). The prevalence of the salmonella

was different between the age of the goats from which the sample was collected. There was significant difference (p < 0.05) in the frequency of Salmonella isolation between adult and young (Table 2).

Table 2: Prevalence of salmonella between the age of goats slaughtered at selected abattoir

Age	Examined	Positive	Prevalence of age%	Prevalence from total%	$X^2$	<i>p</i> -value
Adult	70	1	1.4%	0.5%		
Young	130	4	3.1%	2%	6.4431	0.011
Total	200	5	2.5%	2.5%		

#### Discussions

In the present study the carrier state of salmonella in apparently healthy goats slaughtered at Abyssinia export abattoir was 2.5%. Among the total of 200 animals examined, 5 (2.5%) were positive for Salmonella of which 1 (0.5%) were adult and 4 (2%) were young. The prevalence of Salmonella was higher in young than adult. This difference was statistically significant (P=0.01). This variation in prevalence of Salmonella might be due management differences and handling of owner in this age (Wassie, 2004)<sup>[38]</sup>. The higher prevalence in young might be due to higher Salmonella carrier rate in the study population. The prevalence of Salmonella in apparently healthy slaughtered goats in this study was 2.5%. This result is consistent with the range cited by (D'Aoust, 1989) <sup>[10]</sup> which indicated that the prevalence of Salmonella in goats falls between 1 to 18.8%. also, the result obtained was agreed with the result of Ferede et al., (2015) <sup>[14]</sup> done on the distribution of salmonella was higher in young than adults that was 2%. However, this result is inverse of the previous reports. This variation in result may be due to the young was congregate with your mother and cross contamination during sampling or due to different bacteriological procedures followed. In the present study, Salmonella was isolated in 1.28% of the abdominal muscle of goat. Isolation of salmonella from goat carcass swab was 17.7% in Dire Dawa municipal abattoir in Ethiopia.

The current finding also relatively granted with the reports by Molla, (2005) <sup>[39]</sup>, who detected 0.7% Salmonella isolates, 2% reported by Zubair and Ibrahim (2012) <sup>[41]</sup> and Bedaso *et al.* (2015) <sup>[42]</sup> who reported 0.54% from the slaughtered goats. However, it was lower than the previous findings by Teklu and Negussie (2011) <sup>[35]</sup>, who reported 11.7% in Modjo, Ethiopia and Woldemariam *et al.* (2005) <sup>[40]</sup>, who reported 9.8% in apparently healthy slaughtered sheep and goats in Bishoftu, Ethiopia and other reported by Wassie (2004) <sup>[38]</sup> in Addis Ababa abattoir, which was 3%. The difference in these results might be attributed to the difference in the stress condition and/or exposure of animal to the infected animal due to uncontrolled animal movement while held in the market and lairage which could lead to increased infection rate among the animal. In Zambia Sharma *et al.* (2001) <sup>[43]</sup> was reported prevalence of 2.3% from carcass swab samples of goats. In Ethiopia, Ferede *et al.* (2015) <sup>[14]</sup> which was 17.7% was reported from apparently healthy goats at Dire Dawa municipal abattoir, and 16.7% prevalence reported from goats slaughtered at Elfora abattoir in Ethiopia (Woldemariam, 2003) <sup>[37]</sup>. The current study revealed that as the carcass contaminations of the study area were low and this indicates that they were found at good hygienic condition. The result report variation might be due to the variation in the ecology of study population, sampling technique and procedure sample type and bacteriological techniques based on isolate of Salmonella or distribution of Salmonella in the study population regardless of test samples and methods of detection.

The prevalence of Salmonella on the 40 samples of knife swab was 1(0.5%) at Abyssnia export abattoirs. This result was lower than the results of Teklu and Negussie (2011)<sup>[35]</sup>, who had reported 7.4% Salmonella prevalence from eviscerating knife swabs and other was reported 26.7 and 10% prevalence in two Botswana abattoirs (Motsoela et al., 2002) <sup>[25]</sup>. The variation might be associated with stress during transportation to the slaughterhouse, hygienic conditions of holding pens, processing practices, abattoir facilities and employee's hygiene and practices (Muluneh, 2019) [44]. Usually, healthy carriers' animals intermittently excrete only a few Salmonella, unless they undergo some kind of stress (example during transport or holding in the lairages prior to slaughter). This result was in consistence with Molla et al., (2006)<sup>[24]</sup>. It is generally accepted that the carcass of healthy slaughtered animals are free of bacteria at the time of slaughter, assuming that the animals are not in a state of exhaustion (Jay, 2000) [21]. These differences in prevalence of abdominal muscle may be due to hygienic condition of abattoirs and its environment.

#### **Conclusion and Recommendations**

In the present study, an overall of 2.5% Salmonella was isolated from 200 different types of samples examined from slaughtered goats at Abyssnia export abattoirs. The isolation of this number showed that slaughtering goat are important sources of meat contamination with the organism, and consumption of raw meat can lead to infection with zoonotic salmonellosis.

Therefore, based on the present study and findings, the following points are forwarded:

- Application of good hygiene practice is mandatory during slaughtering processes
- Rational use of veterinary antimicrobials for animal necessary to prevent subclinical diseases
- It should be necessary to provide public education to aware people not to consume raw meat.
- Further investigations and studies of Salmonella up to molecular characterization should be carried out.

### Acknowledgements

First for most Glory to Allah in the highest for helping me in my entire life, without his assistance nothing would be and impossible to be here. I would like to express my deepest gratitude to my advisor Dr. Megarsa Bedasa for his overall guidance and advices, sharing her knowledge from the beginning to the end of my study. I am deeply indebted to my beloved relative Mr. Ephrem Shimalis and my beloved family my father Sadek Mohamed and my mother Naima Aliyi, my brothers Abdi Xaha and Yaken Sadek, and my sisters Alfiya Sadek and Oromia Sadek, for their unconditional love and support all throughout the years, without their unlimited support and encouragement, I wouldn't have been here yet. Your kindness means a lot to me just thank you. I wish thank you my friends, Mustariya Sabur, Cala Haji, Ramadan Nur, Mr. Firomsa Abdalla, Dr. Ibrahim Aliyi, Alemu Aga, Abdulmenan Kader, Wario Boru, Derartu Abdi and Gech Said, for their continuous support and feedbacks before and after I joined university. Once again thanks my lovely friends. Last but not the least appreciation is also given to the following individuals for their positive and helpful assistance throughout my education period: my uncle Abdujabar and Abdusamad Mohamed and especially I want to appreciating Mrs. Bilisuma Abebe for her helpful and supporting me throughout my study and all others. Africa. 98: 26-30.

10. ESGPIP (2007) Body Condition Scoring of Sheep and Goats. Technical

Bulletin p. 8 9. Vatta AF (2005) Goat and Sheep keepers' veterinary manual. Onderstepoort veterinary institute, private bag underreports, South Africa. 98: 26-30.

10. ESGPIP (2007) Body Condition Scoring of Sheep and Goats. Technical

Bulletin

#### Abbreviations

AAU Addis Ababa University CVMA Collage of Veterinary medicine and agriculture IBC Institute of Biodiversity Conservation ISO International Organization for Standardization Masl meter above sea level MR Methyl red OIE Office International Des Epizootics PWB Peptone water broth RVS Rappaport-Vassiliadis soy peptone TSI Triple sugar iron VP Voges Proskauer XLD Xylose lysine deoxycholate

#### Reference

- 1. Domestic animal genetic resources of Ethiopia. IBC (Institute of Biodiversity Conservation), Addis Ababa, Ethiopia; c2007.
- 2. Arruda SGB, Biscontini TMB, Stamford TLM. Microbiological characterization of goat meat subjected to different forms of management. Hygiene Alimentary. 2004;18:58-62.
- Alemayehu D, Molla B, Muckle A. Prevalence and antimicrobial resistance pattern of Salmonella isolates from apparently healthy slaughtered cattle in Ethiopia. Tropical Animal Health and Production. 2003;35:309-319.
- 4. Behnke RH. The contribution of livestock to the economies of IGAD member states: study findings, application of the methodology in Ethiopia and recommendations for further work. IGAD. 2010;10:1-45.
- Blood DC, Henderson AJ, Radosontits AJH, Gay CC. A Text Book of the Diseases of Animals. 9<sup>th</sup> ed.; c2003. p. 809-829.
- 6. Central Statistics Agency. Agricultural sample survey. Livestock and livestock characteristic report (Private

Peasant Holdings). April. Addis Ababa, Ethiopia. 2017;2:1-83.

- 7. Cooke EM, Todd E. Epidemiology of foodborne illness: UK. Lancet. 1990;336:790-793.
- 8. Carlton LD. Salmonellosis: the merk veterinary manual. 8th Edn. NJ; c1998. p. 120-123.
- 9. D'Aoust JY. Salmonella and the international food trade. Int J Food Microbiol. 1994;24:11-31.
- D'Aoust JY. Salmonellae. In: Doyle M (9th Edn) Food borne bacterial pathogens. Marcel Dekker Inc. New York; c1989. p. 328-413.
- Dworkin M, Falkow S, Rosenberg E, Schleifer K, Stackebrandt E. The Prokaryotes: A Handbook on the Biology of Bacteria. 3<sup>rd</sup> edn, 01. 2006, 1.
- Endalew B, Ayalew Z. Assessment of the role of livestock in Ethiopia: A Review. Am Eur J Sci Res. 2016;11:405-410.
- 13. Freeman BA. Burrows Textbook of Microbiology. 22nd edn. W.B. Saunders Company; c1985. p. 464-472.
- 14. Ferede B, Desissa F, Feleke A, Tadesse G, Moje N. Prevalence and antimicrobial susceptibility of Salmonella isolates from apparently healthy slaughtered goats at Dire Dawa municipality abattoir, Ethiopia. J Microbial Antimicrob. 2015;7(1):1-5.
- 15. Gould LH, Mungai E, Behravesh CB. Outbreaks attributed to cheese: differences between outbreaks caused by unpasteurized and pasteurized dairy products, United States, 1998-2011. Foodborne Pathog Dis. 2014;11:545–51.
- Gene O. The isolation, identification and serotyping of Salmonella isolated from domestic poultry in Kars district. Kafka's Univarsikesi Veteriner Fakultesi Dergisi. 2002;8:23-30.
- 17. Ganteby RM. Sheep: The Tropical Agriculturalist. Basingstoke and Macmillan education Limited, London; c1991. p. 1-2.
- House JK, Smith BP. Current strategies for managing salmonellae infection in cattle. Veterinary medicine; c1998. p. 756-764.
- 19. Hansson I, Hamilton C, Ekman T, Forslund K. Carcass quality in certified organic production, compared with conventional livestock production. J Vet Med B Infect Dis Vet Public Health. 2000;47:111-120.
- International Organization for Standardization (ISO). ISO 6579: Microbiology of Food and Animal Feeding Stuff- Horizontal Method for the Detection of Salmonella, (ISO, Geneva); c1998.
- 21. Jay JM. Modern Food Microbiology. 6th ed. Maryland: Aspen Publications; c2000. p. 511-525.
- 22. Kadaka J, Itokazy K, Nakamura M, Taira K, Asato R. An outbreak of Salmonella Weltevreden food poisoning after eating goat meat. Infect Agents Sur Rep. 2000;21:164.
- 23. Molla B, Mohammed A, Salah W. Salmonella prevalence and distribution of serotypes in apparently healthy slaughtered camels (Camelus dromedarius) in Eastern Ethiopia. Trop Anim Health Prod. 2004;36(5):451–458.
- 24. Molla W, Molla B, Alemayehu D, Muckle A, Cole L, Wilkie E. Occurrence and antimicrobial resistance of Salmonella serovars in apparently healthy slaughtered sheep and goats of central Ethiopia. Trop Anim Health Prod. 2006;38:455-462.

- Motsoela C, Collison EK, Gashe BA. Prevalence of salmonella in two Botswana abattoir environments. J Food Prot. 2002;65:1869-1872.
- Office International Des Epizootics (OIE). Manual of Standards for Diagnostic Tests and Vaccines; c2000. p. 37.
- Pieskus J, Milius J, Michalskiene I, Zagrebneviene G. He distribution of Salmonella serovars in chicken and humans in Lithuania. J Vet Med A Physiol Pathol Clin Med. 2006;53:12-16.
- Pepin M, Russo P, Pardon P. Public health hazards from small ruminant meat products in Europe. Rev Sci Technol. 1997;16(2):415-425.
- Quinn PJ, Carter ME, Markey BK, Carter CR. Veterinary Clinical Microbiology. Wolfe Publishing; c1994.
- 30. Quinn PJ, Markey BK. Concise review of veterinary microbiology. Backwell Publishing; c2003. p. 6.
- Rabsch W, Altier C, Tschape H, Baumler AJ. Food borne Salmonella infection. In: Torrence ME, Isaacson RE, editors. Microbial food safety in animal agriculture. Blackwell Publishing; c2003. p. 97-108.
- Reid C, Small A, Avery S, Buncic S. Presence of foodborne pathogens on cattle hides. Food Control. 2002;13:411-415.
- 33. Sierra M, Gonzales-Fandos E, Garcia-Lopez M, Fernandez M, Prieto M. Prevalence of Salmonella, Yersinia, Aeromonas, Campylobacter and cold-growing Escherichia coli on freshly dressed lamb carcasses. J Food Prot. 1995;58:1183-1185.
- Tabaraie B, Sharma BK, Sharma PR, Sehgal NR, Ganguly NK. Evaluation of Salmonella porins as a broad-spectrum vaccine candidate. Microbio. Immuno. 1994;38:553-559.
- 35. Teklu A, Negussie H. Assessment of risk factors and prevalence of Salmonella in slaughtered small ruminants and environment in an export abattoir, Modjo, Ethiopia. Am Eurasian J Agric Environ Sci. 2011;10(6):992-999.
- 36. Vatta AF. Goat and Sheep keepers' veterinary manual. Onderstepoort Veterinary Institute; 2005. p. 26-30.
- Woldemariam E. Prevalence and distribution of Salmonella in apparently healthy slaughtered sheep and goats in Debre Zeit, Ethiopia. DVM Thesis. AAU, FVM, Debre Zeit, Ethiopia; c2003.
- 38. Wassie M. A cross-sectional study on Salmonella in apparently healthy slaughtered sheep and goats at Addis Ababa and Modjo abattoirs, Ethiopia. MSc Thesis. Addis Ababa University, Faculty of Veterinary Medicine, Bishoftu, Ethiopia; c2004.
- 39. Molla W, Molla B, Alemayehu D, Muckle A, Cole L, Wilkie E. Prevalence and distribution of Salmonella in apparently healthy slaughtered Occurrence and antimicrobial resistance of Salmonella serovars in apparently healthy slaughtered sheep and goats of central Ethiopia. April. [Internet]; c2005. Available from: https://doi.org/10.1007/s11250-006-4325-4
- 40. Woldemariam E, Molla D, Alemayehu D, Muckle A. Prevalence and distribution of salmonella in apparently healthy slaughtered sheep and goats in Debrezeit, Ethiopia. Small Rumin Res. 2005;58:19-24.
- 41. Zubair AI, Ibrahim KS. Isolation of Salmonella from slaughtered animals and sewage at Zakho abattoir,

Kurdistan Region, Iraq. Res Opin Anim Vet Sci. 2012;3(1):20-24.

- 42. Bedaso KK, Dinsefa JH, Kifle AL, Biniam T. Study on prevalence and distribution of Salmonella isolates from apparently healthy sheep and goats slaughtered at Addis Ababa Abattoir enterprise, Ethiopia. Journal of Veterinary Science and Technology. 2015, 6(6).
- 43. Sharma R, Chisti Y, Banerjee UC. Production, purification, characterization, and applications of lipases. Biotechnology advances. 2001 Dec 1;19(8):627-62.
- 44. Woday A, Muluneh A, St Denis C. Birth asphyxia and its associated factors among newborns in public hospital, northeast Amhara, Ethiopia. PloS one. 2019 Dec 20;14(12):e0226891.